

[7]. Further, P-III SVMPs are major components of rear-fanged snake venoms and account for several of the effects observed following envenomation [11,12]. However, there are few studies on the primary structure of P-III SVMPs found in rear-fanged snake venoms. Moreover, there are no published studies on the autocatalytic processing, either in vivo or in vitro. The venom metalloproteinase is a 57.5-kDa metalloproteinase isolated from the *Philodryas patagoniensis* (Patagonia Green Racer), a South American rear-fanged snake which is now considered a member of the *Philodryadidae* [13]

enzyme and its main autoproteolytic fragment, we demonstrated that it is subjected to specific autoproteolytic cleavage at the initiation of the disintegrin domain.

(data not shown), indicating that it is still connected by disulfide

band of ~32.6 kDa (Fig. 1B), whose molecular mass is compatible with disintegrin-like/cysteine-rich domains [18]. This fragment is not observed when non-reduced samples were submitted to SDS-PAGE



Fig. 1. Sequence alignment of the N-terminus of the 32.6-kDa autoproteolytic fragment of patagonfibrase (red), and peptides sequenced by LC-MS/MS (see Table 1) from patagonfibrase and its 32.6-kDa autoproteolytic fragment, with other P-III SVMPs. Protein sequences were aligned using the program ClustalW [34]. The numbers indicate the amino acid residues of a metalloprotease from *P. olfersii*, POLF0061C [11]. The other SVMPs were referenced by their GenBank accession numbers: ACS74988, from *P. olfersii* (Dipsadidae); ABU68535, from *Thrasops jacksoni* (Colubridae); P82942, kaouthiagin, from *Naja kaouthia* (Elapidae); CAA48323, jararhagin, from *Bothrops jararaca* (Viperidae); and CAJ01683, from *Echis ocellatus* (Viperidae). Identical residues are boxed in black. The boundary between the metalloproteinase and disintegrin domains of SVMPs is indicated, and disintegrin-like sequences are shown in green.

parent structure play an important role in stabilizing and tightening the segment connecting the proteolytic domain with the succeeding disintegrin domain [19,22]. As shown here, patagonfibrase structure is partially stabilized by Ca^{2+} , since there is a proteolysis product at 52.2 kDa in the presence of this ion. This fragment likely represents patagonfibrase processed to release a portion of the amino-terminal

- [16] H. Blum, H. Beier, H.J. Gross, Improved silver staining of plant proteins, RNA and DNA in polyacrylamide gels, *Electrophoresis* 8 (1987) 93–99.
- [17] S.L. Hanna, N.E. Sherman, M.T. Kinter, J.B. Goldberg, Comparison of proteins expressed by *Pseudomonas aeruginosa* strains representing initial and chronic isolates from a cystic fibrosis patient: an analysis by 2-D gel electrophoresis and capillary column liquid chromatography-tandem mass spectrometry, *Microbiology* 146 (Pt 10) (2000) 2495–2508.
- [18] J. Zang, Z. Zhu, Y. Yu, M. Teng, L. Niu, Q. Huang, Q. Liu, Q. Hao, Purification, partial characterization and crystallization of acucetin, a protein containing both disintegrin-like and cysteine-rich domains released by auto-proteolysis of a P-III-type metalloproteinase AaH-IV from